GENTIOLACTONE, A SECOIRIDOID DILACTONE FROM GENTIANA PURPUREA

INGE H. SUHR, PETER ARENDS and BIRTHE JENSEN

Department of Chemistry BC, Royal Danish School of Pharmacy, 2 Universitetsparken, DK-2100 Copenhagen Ø, Denmark (Received 20 June 1977)

Key Word Index—Gentiana purpurea; Gentianaceae; secoiridoid; gentiolactone.

Abstract—The structure of gentiolactone, a new secoiridoid dilactone isolated from the dried roots of *Gentiana purpurea* was elucidated by spectroscopic, spectrometric and X-ray diffraction methods as (\pm) -4-ethyl-4-hydroxy-3,4,5,6-tetrahydro-1H,8H-pyrano[3,4-c]-pyran-3,8-dione.

INTRODUCTION

The isolation and characterization of several constituents of *Gentiana purpurea* have been reported [1-7]. During a search for phenolic compounds [8] in root material of this species, a compound which later proved to be a secoiridoid was isolated and its structure (1) elucidated by spectroscopic and spectrometric methods. The compound shows no optical activity and an X-ray diffraction analysis was undertaken to confirm the structure and show that the compound is racemic.

RESULTS AND DISCUSSION

In the MS of 1 a very small peak at m/e 212 was found to represent the parent ion for both the m/e 183 ($C_8H_7O_5^+$) and m/e 168 ($C_9H_{12}O_3^+$) ions, thus it was a reasonable candidate for M^+ . Its composition ($C_{10}H_{12}O_5^+$) was deduced from its fragmentations which proceed through loss of CO_2 and $C_2H_5^+$ by alternative routes leading to the base peak ion at m/e 139 ($C_7H_7O_3^+$).

The IR spectrum of 1 in KBr showed a remarkably broad and intense carbonyl band at 1725–1715 cm⁻¹ which recorded in CHCl₃ was split to give 1725 and 1745 cm⁻¹ bands. In conjunction with prominent bands in the C—O stretching region and the loss of CO₂ in the MS the presence of lactone groups were confirmed. Bands close to 3500 cm⁻¹ (OH), and a weak band at 1600 cm⁻¹

in the solution spectrum (C=C conjug.) also fit the structure.

In the PMR spectrum of 1 in CDCl₃, signals at δ 1.01 (3H, t) and at δ 1.5–2.1 (2H, m) were found by double resonance experiments to represent an ethyl group which was not further proton coupled. The non-equivalence of the methylene protons, which may be approximated as mutually displaced quartets was in agreement with positioning of the ethyl group at C-4, a chiral centre [9]. The presence of the hydroxyl group was confirmed by a signal at δ 3.6 (1H, s) which disappeared upon addition of D₂O. In DMSO-d₆ this signal was at δ 6.15 and was still a singlet indicating that the alcoholic function was tertiary [10].

The remaining signals in the PMR spectrum represented the ring methylene groups. A signal at δ 4.54 was a triplet (J = 6.5 Hz) while at δ 4.9-5.5 was seen an AB system of doublets (J = 16.5 Hz), each further split into triplets (J = 2.5 and 1.5). By irradiation at the frequency of a signal at δ 2.6-2.9 (m) the methylene triplet collapsed into a singlet while the triplet splitting of the other methylene protons disappeared. The low-field protons were those at C-6 and C-1, deshielded by the neighbouring oxygen atoms. The latter set were further deshielded by the double bond and are obviously magnetically non-equivalent. Their homoallylic coupling with the methylene protons at C-5 have constants of expected magnitude for the transoid case [11].

In the 13 C-NMR spectrum of 1 in CDCl₃ signals at δ 7.7 (q) and 22.7 (t) corresponded to the ethyl group. A signal at δ 31.0 (t) arose from the allylic methylene carbon C-5. A signal at δ 66.7 (t) corresponded to the methylene carbon atoms bonded to oxygen in the ester groupings; remarkably, they were isochronous although their protons were not. The carbonyl functions showed up at δ 172.6(s) and 161.6(s), in good agreement with the values expected for a six-membered lactone (ca δ 175) and for its α , β -unsaturated counterpart, since the latter may be expected to resonate at 5–15 ppm higher field in analogy with other carbonyl compounds [12]. The α and β carbon signals were seen at δ 120.0 (s) and 153.4 (s), respectively. A signal at δ 72.3 (s) was assigned to the carbon atom carrying the alcohol group.

The UV absorption was consistent with the chromo-

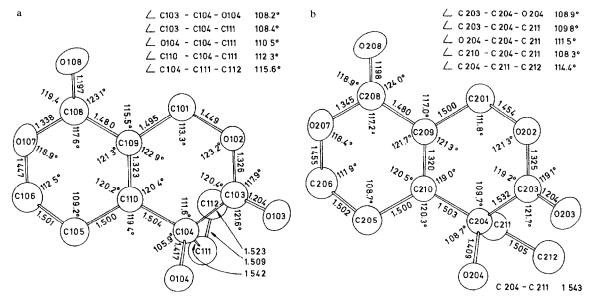


Fig. 1a and b. Bond lengths (Å), angles (°) and conformations of the two crystallographically independent molecules of gentiolactone. The estimated standard deviations on bond lengths and angles are 0.004–0.006 Å and 0.2–0.3, respectively. The drawings were produced by ORTEP II [17].

phore present in 1. α,β -Unsaturated acids and esters in which the double bond is fully substituted absorb at $225 \pm 5 \text{ nm}$ [13] and the same may be expected to hold true for lactones.

Table 1. Final positional parameters

Atom	x/A	y/\mathbf{B}	z/\mathbf{C}
C101	0.3296 (4)	0.9588 (3)	0.0729 (3)
O102	0.4586(3)	0.9722(2)	0.1598(2)
C103	0.4409 (4)	0.9831(3)	0.2836(3)
C104	0.2799 (4)	1.0011(3)	0.3360(3)
C105	0.0041 (4)	0.9011(3)	0.2977(3)
C106	-0.1207(5)	0.9043 (4)	0.2014 (4)
O107	~ 0.0773 (3)	0.8357(3)	0.0833(2)
C108	0.0648 (4)	0.8549(3)	0.0413(3)
C109	0.1801(4)	0.9235(3)	0.1282(3)
C110	0.1558 (4)	0 9412 (3)	0.2418(3)
O103	0.5557(3)	0.9828(3)	0.3442(2)
O104	0.2739(3)	0.9327(3)	0.4392(2)
O108	0.0944(3)	0.8138(3)	-0.0632(2)
C111	0.2543 (4)	1.1475 (3)	0 3756 (4)
C112	0 2616 (4)	1 2316 (4)	0 2739 (3)
C201	0.8008(4)	0.5306(3)	0.0875(3)
O202	0.9337(3)	0.5353(3)	0.1710(2)
C203	0.9167 (4)	0.5228(3)	0.2887(3)
C204	0.7544 (4)	0.4974(3)	0.3361(3)
C205	0.4899 (4)	0.6172(3)	0 3330 (3)
C206	0.3608 (4)	0.6112 (4)	0.2389 (4)
O207	0.4039(3)	0.6743(3)	0.1336(2)
C208	0.5417 (4)	0.6452(3)	0.0813(3)
C209	0.6566 (4)	0.5798(3)	0.1526(3)
C210	0 6358 (4)	0.5688(3)	0.2697(3)
O203	1.0293 (3)	0.5320(3)	0.3557(2)
O204	0.7498(3)	0 5443 (2)	0.4626(2)
O208	0 5659 (3)	0.6752(3)	-0.0185(2)
C211	0.7161(4)	0 3508 (3)	0.3112(3)
C212	0.8278 (5)	0.2661 (4)	0.3701 (4)
H104	0.338 (5)	0.961 (4)	0.488(4)
H204	0.836 (5)	0.523 (4)	0.498(3)

The X-ray diffraction analysis showed that the compound was racemic. The final positional parameters are listed in Table 1. The molecular dimensions and conformations of the two crystallographically independent molecules are given in Fig. 1a and b together with the numbering used for the non-hydrogen atoms. The one major difference between the two independent molecules was found for the torsion angle Cx10–Cx04–Cx11–Cx12, the value of which was $\pm 63.4(4)^{\circ}$ for x = 1 and ± 178.8 (3)° for x = 2. The crystal packing is illustrated in Fig. 2. Inter-molecular hydrogen bonds Ox04-Hx04...Ox03 link molecules related by a center of symmetry together in pairs. The dimensions of the hydrogen bonds are listed in Table 2 together with distances and angles for the intra-molecular contacts Ox04-Hx04...Ox03. The crystal structure is stabilized by a great number of van der Waals' contacts.

The structure of gentiolactone includes the secoiridoid carbon skeleton, a taxonomic feature already associated with the Gentianaceae [14]. Gentiopicroside (2) a member of the same class is found in roots of G. purpurea [5]. The possibility that gentiolactone is formed from this compound may be considered. In that case the configurational identity of the side chain-bearing carbon atom is lost in the process.

Table 2. Inter-molecular hydrogen bonds and intra-molecular contacts Ox04-Hx04. Ox03. Distances (Å) and angles (°)

A-H B	A-H	A B	H B	∠AHB
O104-H104 O103 _(1-x, 2-y, 1-z)	0.79 (4)	2.810 (4)	2.05 (4)	159 (4)
O204-H204 O203 _(2-x 1-y,1-z) O104-H104O103 O204-H204O203	0.79 (4)	2.927 (4) 2.728 (4) 2.688 (4)	2.49 (4)	154 (4) 99 (3) 107 (3)

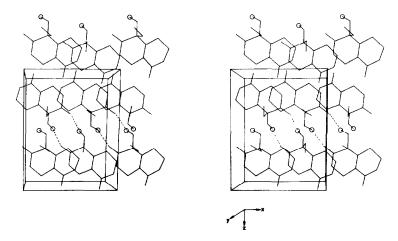


Fig. 2. Stereo view of the crystal packing. The hydroxyl hydrogen atoms are indicated by circles and hydrogen bonds by broken lines.

EXPERIMENTAL

Extraction and purification. Dried roots of Gentiana purpurea L. were available commercially from Caesar & Loretz, Hamburg. The powdered root material (1 kg) was defatted with petrol, then extracted with CH₂Cl₂ (Soxhlet) for about 70 hr. Evaporation afforded 15.8 g residue which was taken up in 90% MeOH. The soln was extracted with petrol and the MeOH phase evapd. Column chromatography on Si gel (700 g) in CH₂Cl₂ afforded 1 in the fractions eluted with CH₂Cl₂-EtOAc (99:1). Further purification was effected on a similar column eluted with C₆H₆-EtOAc mixtures, yielding 180 mg compound 1. Crystallization from EtOH afforded colourless irregular prisms, mp 119–120.5° (corr.). TLC on Si gel F_{254} , CHCl₃–EtOAc (4:1) showed a single spot, R_f 0.21 in short wave UV light. With the NH₂OH-FeCl₃ spray reagent a weak colour reaction was also obtained. TLC on polyamide $F_{2.54}$ (Merck Fertigplatten) using MeOH also showed a single spot, R_f 0.75. $[\alpha]_{\lambda}^{25}$ 0° (MeOH; 0.855) for λ between 365 and 589 nm. $\lambda_{\rm max}^{\rm MeOH}$ nm $\log \varepsilon$): 225–230 (3.74), decreasing with time upon addition of NaOMe, unaffected by HCl. $\lambda_{\max}^{\text{MeCN}}$ nm (log ε): 227 (3.75). ν_{\max}^{KBr} cm⁻¹: 3490 s, 3460 s, 3020 w, 2980 w, 2940 w, 2920 w, 2850 w, 1725-1715 vs, 1470 m, 1445 w, 1425 w, 1410 m, 1375 w, 1350 w, 1325 m, 1310 s, 1295 w, 1260 m, 1230 w, 1220 w, 1205 m, 1180 m, 1170 m, 1155 s, 1135 s, 1110 m, 1065 m, 1040 m, 1020 s, 995 m, 965 w, 940 w, 920 w, 890 w, 815 w, 760 m, 720 w, 660 w, 625 w.

MS. (AEI-MS 902 instrument, probe) 70 eV, m/e (rel. int.): 212 (0.5), 184 (8), 183 (37), 182 (7), 169 (7), 168 (40), 156 (3), 155 (4), 150 (3), 140 (16), 139 (100), 138 (9), 137 (3), 125 (4), 124 (3), 123 (16), 122 (4), 121 (32), 112 (3), 111 (7), 109 (4), 108 (2), 98 (2), 97 (4), 95 (6), 93 (4), 91 (4), 83 (5), 82 (3), 81 (12), 80 (4), 79 (5), 77 (5), 69 (7), 68 (5), 67 (37), 66 (7), 65 (14), 63 (3), 57 (48), 56 (3), 55 (8), 53 (19), 52 (12), 51 (12), 50 (5). m^* , 134.0, 115.0 (168 \rightarrow 139), 105.5 (183 \rightarrow 139), 91.0, 90.1, 88.8, 84.7 and 63.1. Metastable defocusing: 212 \rightarrow 168, 212 \rightarrow 183 and 196 \rightarrow 168. High resolution measurements: 183.0293, 168.0786 and 139.0395.

NMR. PMR spectra were recorded on a Bruker HX-90-E instrument. The ¹³C NMR spectrum was recorded on a Bruker WH-90 instrument. Chemical shifts are relative to TMS as the internal standard.

X-ray analysis. The material used for the X-ray examination was recrystallized from CHCl₃ and single crystals were grown

from a mixture of CHCl₃ and CCl₄ by slow evaporation at 5°.

Crystal data. 4-Ethyl-4-hydroxy-3,4,5,6-tetrahydro-1H,8H-pyrano[3,4-c]pyran-3,8-dione, $C_{10}H_{12}O_5$, M=212.20. Triclinic, a=8.614 (3), b=10.382 (3), c=10.973 (2) Å, $\alpha=97.28$ (2), $\beta=90.76$ (3), $\gamma=90.35$ (3)°. U=972.2 ų. D_m (flotation) = 1.448 g cm⁻³, Z=4, $D_c=1.450$ g cm⁻³. Linear absorption coefficient for X-rays [λ (MoK $\alpha=0.7107$ Å], $\mu=1.25$ cm⁻¹, F(600)=448. Space group $P\bar{1}$. Three-dimensional diffraction data were measured at room temp. on a Nonius three-circle automatic diffractometer using standard techniques. The structure was solved and refined using MULTAN [15] and programs of the X-RAY-system [16], respectively. Detailed information about experimental conditions and the refinements are, together with the final lists of structure factors, thermal parameters and calculated positional parameters for hydrogen atoms, available from the authors on request.

Acknowledgements—One of us, Inge H. Suhr, thanks The Royal Danish School of Pharmacy for a research scholarship.

REFERENCES

- Hegnauer, R. (1966) Chemotaxonomie der Pflanzen, band 4, p. 176. Birkhäuser Verlag Basel, and references cited therein.
- Luckner, M., Bessler, O. and Schröder, P. (1965) Pharmazie 20, 16
- 3. Franz, G. (1971) Verhandl. Schweiz. Naturforsch. Ges. 151, 75.
- Verney, A.-M. and Debelmas, A.-M. (1973) Ann. Pharm. Franc. 31, 415.
- Wagner, H. and Vasirian, K. (1974) Deut. Apotheker Ztg. 114, 1245.
- Hostettmann, K. and Jacot-Guillarmod, A. (1974) Phytochemistry 13, 1625.
- 7. Hostettmann, K., Duc, L. M., Goetz, M. and Jacot-Guillarmod, A. (1975) *Phytochemistry* 14, 499.
- 8. Suhr, I. H. Unpublished results.
- Bishop, E. O. (1961) Ann. Rept. Progr. Chem. (Chem. Soc London) 58, 55.
- Chapman, O. L. and King, R. W. (1964) Chem. Commun. 86, 1256

- Jackman, L. M. and Sternhell, S. (1969) Applications of Nuclear Magnetic Resonance Spectroscopy in Organic Chemistry, 2nd Edn, p. 325 Pergamon Press, London.
- Levy, G. C. and Nelson, G. L. (1972) Carbon-13 Nuclear Magnetic Resonance for Organic Chemists p. 119. Wiley Interscience, New York.
- 13. Nielsen, A. T. (1957) J. Org Chem 22, 1539.
- 14. Inouye, H. (1970) Pharmacognosy and Phytochemistry. 1. International Congress, Munich (Wagner, H. and
- Hörhammer, L. eds) p. 290. Springer, Berlin (1971).
- 15 Declercq, J. P., Germain, G., Main, P. and Woolfson, M. M (1973) Acta Cryst. A29, 231.
- Stewart, J. M., Kruger, G. J., Ammon, H. L., Dickinson, C. H. and Hall, S. R. X-RAY 72, Tech. Rep. TR 192, Computer Science Center, Univ. of Maryland, College Park, Maryland.
- 17. Johnson, C. K. ORTEP II. Oak Ridge National Laboratory Report ORNL-3794, revised 1971.